

BREEDING FOR SUSTAINABILITY: HOW REPRODUCTIVE BIOTECHNOLOGIES CAN HELP BUFFALO FARMERS COMBAT CLIMATE CHANGE

Reproduciendo para la sostenibilidad: cómo las biotecnologías reproductivas pueden ayudar a los criadores de búfalos a combatir el cambio climático

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ABSTRACT

The global attention on enteric CH₄ production in ruminants requires a response that involves collaboration between researchers and industry. Future generations of buffaloes will be characterized by better efficiency and fertility, which may reduce CH₄ emission intensity. This goal will result from balanced multi-trait selection and the introduction of efficient reproductive and productive management. Currently, efficient reproductive programs using assisted reproductive technologies (ARTs) are available on buffalo farms. Our expanding knowledge of ovarian function during the buffalo estrous cycle has given new approaches for precisely synchronizing follicular development and ovulation to apply ARTs consistently. Synchronization protocols are designed to control both luteal and follicular function and permit fixed-time AI with high pregnancy rates during the breeding (autumn-winter) and non-breeding (spring-summer) seasons. Additionally, it allows the initiation of superstimulatory treatments at a self-appointed time, providing opportunities to superstimulate buffalo donors associated with ovum pick-up (OPU) and *in vitro* embryo production (IVEP). Furthermore, it allows fixed-time embryo transfer in recipients, with high efficiency and no need for estrus detection. Thus, ARTs, such as AI and ET, are applied for buffalo's targeted multiplication and dispersal with defined production and environmental credentials. Also, the urgency in moving to the next generation of buffaloes will increase the production of embryos from genomically defined prepubertal heifers. Using these biotechnologies will reduce generation interval and accelerate the rate

of genetic improvement to buffalo, defined by better efficiency and fertility and lower CH₄ emission. The challenge remains to communicate the importance of buffaloes for food security and the environment.

Keywords: enteric methane, efficiency, fertility, assisted reproductive technology.

RESUMEN

La atención mundial sobre la producción de CH₄ entérico en rumiantes requiere una respuesta que implique la colaboración entre investigadores y la industria. Las generaciones futuras de búfalos se caracterizarán por una mayor eficiencia y fertilidad, lo que puede reducir la intensidad de las emisiones de CH₄. Este objetivo será el resultado de una selección equilibrada de múltiples rasgos y la introducción de un manejo reproductivo y productivo eficiente. Actualmente, las granjas de búfalos cuentan con programas reproductivos eficientes que utilizan tecnologías de reproducción asistida (ART). Nuestro creciente conocimiento sobre la función ovárica durante el ciclo estral de las búfalas ha brindado nuevos enfoques para sincronizar con precisión el desarrollo folicular y la ovulación para aplicar las ART de manera consistente. Los protocolos de sincronización están diseñados para controlar la función lútea y folicular y permitir la IA a tiempo fijo (IATF) con altas tasas de preñez durante las temporadas de reproducción (otoño-invierno) y no reproductiva (primavera-verano). Además, permite el inicio de tratamientos de superestimulación en el momento que usted

elija, brindando oportunidades para superestimular a los donantes de búfalos asociados con la recogida de óvulos (OPU) y la producción de embriones *in vitro* (IVEP). Además, permite la transferencia de embriones a tiempo fijo (TETF) en las receptoras, con alta eficiencia y sin necesidad de detección de estro. Por lo tanto, las ART, como la IA y la ET, se aplican para la multiplicación y dispersión selectiva del búfalo con credenciales ambientales y de producción definidas. Además, la urgencia de pasar a la próxima generación de búfalos aumentará la producción de embriones a partir de novillas prepúberes genómicamente definidas. El uso de estas biotecnologías reducirá el intervalo generacional y acelerará la tasa de mejora genética del búfalo, definida por una mayor eficiencia y fertilidad y una menor emisión de CH₄. El desafío sigue siendo comunicar la importancia de los búfalos para la seguridad alimentaria y el medio ambiente.

Palabras clave: metano entérico, eficiencia, fertilidad, tecnología de reproducción asistida.

INTRODUCTION

The world's population is projected to increase by 24% by 2050, potentially reaching 9.7 billion people [1]. Food production must increase by 49% to sustain this population explosion [2]. In this scenario, urbanization, and growing concerns about the environmental impact of livestock farming demand a long-term global strategy for more sustainable ruminant production. Buffalo, therefore, will continue to have a significant role in future global food security. The global buffalo population is approximately 202 million head [3], compared to 1.5 billion cattle [4].

Buffalo milk and meat products can meet human needs for high-quality protein. They excel over cattle exploiting low-quality feed typical of many rearing areas and demonstrate great adaptability to various management and temperature conditions [5]. Furthermore, most buffalo production is carried out extensively in pastures and savannas suited for low-input and low-cost animal production. In South Asia, the River buffalo is a primary source of milk and meat and has a crucial role in food security. The riverine buffalo also supports high-value, differentiated food production in Europe and the Americas. The Swamp buffalo is a vital draft animal and a source of food in Southeast Asia and East Asia.

However, the environmental impact of ruminant production has gained significant attention worldwide [6, 7, 8]. Cattle contribute around 4.5-5.0% of global anthropogenic methane [9]. Enteric fermentation, with an annual emission of 87-97 Tg (i.e., 1012g), is one of the agricultural sector's significant methane sources [10]. The global contribution of cattle and buffalo to annual enteric methane emissions is 77% and 13%, respectively [11]. The primary source of methane in ruminants originates from the enteric fermentation process, where complex carbohydrates are converted into simple sugars by methano-

genic protozoa [12]. Extensive reviews have comprehensively covered the biology and function of the rumen [13, 14]. The quantity of methane an animal produces is significantly influenced by the relative abundance of ruminal methanogenic and non-methanogenic microbes [12]. Microbial gene abundance analysis advancements allow for determining ruminal microbe populations [15]. In addition to enteric methane (CH₄) produced by the rumen, beef, and dairy production also contributes carbon dioxide (CO₂; feed), nitrous oxide (N₂O; feed production, manure), and other CH₄ (manure) to the total greenhouse gas (GHG) budget of the production systems.

Malik et al. [16] compared the enteric methane yield between cattle and buffaloes under the same nutritional management. Enteric methane emissions (g/d) depended on dry matter intake (kg/d). However, the methane yield (g/kg dry matter intake; DMI) did not differ between species when fed on the same diet (Cattle=13.4 g/kg DMI vs. Buffaloes=13.5 g/kg DMI). This result confirms that methane yield depends on the diet rather than the species compared. Thus, methane mitigation strategies developed in one of the species can be effective in the other.

In this scenario, the use of assisted reproductive technologies can have a significant impact on improving efficiency in buffalo production systems. Reproductive technology has been progressively refined in buffaloes, and today, the success of artificial insemination and embryo transfer is comparable to cattle. Artificial insemination (AI), combined with estrus synchronization, is a potent strategy of assisted reproduction technology to improve reproductive efficiency and expedite genetic gain in buffaloes [16]. Furthermore, embryo transfer (ET) enables the multiplication of high maternal and paternal genetic value, playing a more significant role in the genetic enhancement of this species [17]. This review seeks to demonstrate how assisted reproductive technologies (ARTs) can improve reproductive efficiency and harvest the next generation of buffaloes that produce more milk and meat to combat climate change.

REPRODUCTIVE EFFICIENCY IN BUFFALO AND APPLICATION OF ARTIFICIAL INSEMINATION TO IMPROVE PRODUCTION AND REDUCE METHANE EMISSION

The cow-calf operation system utilizes approximately 70% of resources. Therefore, selection for reproductive efficiency significantly affects farm efficiency, profitability, and sustainability. With high reproductive efficiency, fewer cows are required to produce the next generation of calves, reducing resource requirements, herd methane production, and costs [18]. Furthermore, assisted reproduction technologies can also be used to manipulate reproduction in buffalo. This includes synchronization of the breeding time, influencing the age at first breeding, the interval between the calving, and improving the breeding during seasonal anestrus [16].

Artificial insemination can be incorporated into buffalo breeding programs to further improve reproductive efficiency and genetic gain, collaborating to reduce CO₂-eq emission intensity. However, the traditional AI program efficiency needs to be improved by low estrous detection. Buffalo presents a poor manifestation of estrus symptoms, implying operational difficulties in detecting estrus [19].

Furthermore, the success of reproductive programs is closely related to the buffalo reproductive seasonality. Buffalo is a seasonal reproductive species and becomes sexually active in response to a decreasing day length (short days) in late summer to early autumn [20, 21]. During the non-breeding season, buffalo often exhibit anestrus, which extends the anovulatory period and reduces reproductive performance [22].

Nowadays, timed artificial insemination (TAI) can be applied routinely in farm reproductive programs. TAI protocols are designed to control both luteal and follicular function, permitting the AI without estrus detection and during the anestrous period with high reproductive efficiency during the breeding and non-breeding season [19, 23, 24]. Several studies demonstrate that it is possible to establish an effective AI program in buffaloes throughout the year, collaborating to increase the number of pregnant buffaloes during the non-breeding season and distributing calving and milk production throughout the year. Using reproductive programs with TAI followed by resynchronization,

it is possible to obtain high reproductive efficiency (>80% pregnancy rate after 3 FTAI) with inter-calving intervals close to 12 months (FIG. 1; adapted from Baruselli et al. [25]).

The efficiency of TAI in buffalo demonstrates that it is possible to introduce efficient artificial insemination programs on farms that collaborate to increase the reproductive and genetic efficiency of the herds.

EMBRYO TECHNOLOGY TO MITIGATE METHANE EMISSION

In vivo, (superovulation; SOV) and *in vitro* (ovum pick-up and *in vitro* embryo production; OPU/IVEP) embryo productions are reproductive biotechnologies used worldwide in beef and dairy operations to disseminate the genetic material of superior animals. Selection and genetic gain are essential to improve efficiency, product quality, and sustainability [16]. When comparing both biotechnologies in buffalo, OPU/IVEP demonstrates higher efficiency and greater commercial applicability than SOV. However, there are some limitations to using OPU/IVEP, such as seasonality, the low number of antral follicles, and the low quantity and quality of the recovered oocytes [17].

Experiments have been conducted to enhance OPU/IVEP efficiency. In one study, Sá Filho et al. [26] demonstrated

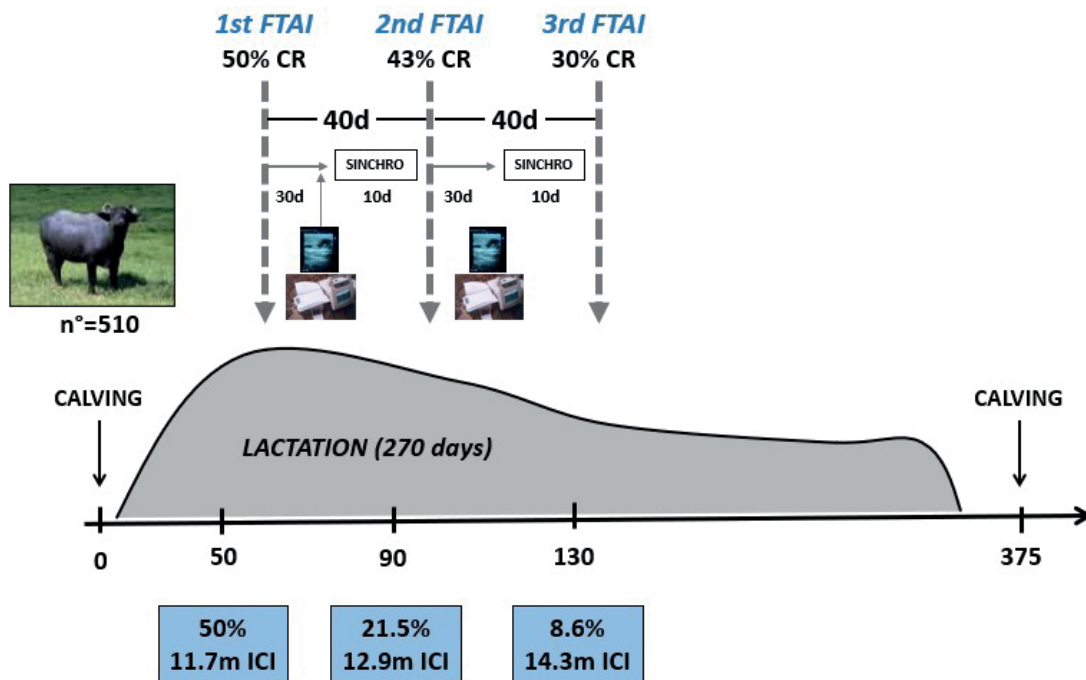


FIGURE 1. Conception rate (P/AI) of lactating buffalo (n=510) submitted to fixed time AI following resynchronization in non-pregnant cows. Ultrasonography evaluation was performed to detect non-pregnant buffaloes 30 days after AI for re-synchronization. Pregnant buffaloes from the 1st FTAI with 50 days postpartum presented a 50% conception rate (CR) and 11.7 months of inter-calving interval (ICI). Pregnant buffaloes from the 2nd FTAI with 90 days postpartum presented a 43% CR and 12.9 months of ICI. Pregnant buffaloes from the 3rd FTAI with 130 days postpartum presented a 30% CR and 14.3 months of ICI. After 3 FTAI, buffaloes presented an 80.1% pregnancy rate with a mean of 12.3 months of ICI (adapted from Baruselli et al., 2003)

that using bST increased the number of small antral follicles at OPU. Additionally, bST tended to increase the number of recovered oocytes and improved the percentage of high-quality oocytes. However, bST showed no effect on cleavage and blastocyst production rates. In another study, Carvalho et al. [27] showed that FSH treatment for superstimulation before aspiration improved the outcomes of OPU/IVEP. FSH treatment increased the proportion of large and medium follicles at OPU and enhanced the viable oocyte rate, blastocyst rate, and number of embryos produced per OPU session.

The use of OPU/IVEP in females before puberty, apart from the genetic gain inherent in this biotechnology, also reduces the generation intervals, further accelerating genetic improvement. This technology can be employed in prepubertal buffalo heifers, where ovaries have established follicular waves and respond to superstimulation, or in buffalo calves, where OPU is performed via laparoscopy [LOPU; 28, 29, 30]. LOPU permits the recovery of oocytes from calves of two months of age and the *in vitro* production of embryos that will be transferred to recipients. This technology allows a donor animal to produce offspring before it reaches sexual maturity. The use of young donors has two main key points that make this alternative interesting: the first one is the larger follicular population, and the number of cumulus-oocyte complexes (COCs) recovered, and the second is the shorter generation interval, increasing genetic gain [30, 31].

In a study conducted by our group, we compared embryo production in buffalo calves (2-4 months of age), prepubertal buffalo heifers (13-15 months of age), and lactating buffalo cows [28]. The treatment for calves involved using a sheep intravaginal P4 device on day 0 of the protocol, and for stimulating follicular growth, 140 mg of FSH was administered in four decreasing doses every 12 hours on days 5 and 6. On day 7, oocytes were recovered by LOPU in calves and through OPU on a random day of the estrous cycle in prepubertal heifers and lactating cows. The results showed that calves had a lower blastocyst production rate, but the number of embryos produced was similar between calves and lactating cows. Embryos produced from calves ($n=8$) resulted in three pregnancies (3/8; 38%), which led to the birth of three healthy calves [28]. This study demonstrated the feasibility of IVEP in young animals to reduce generation interval and significantly accelerate genetic progress in buffaloes. However, calves were less efficient in embryo production than prepubertal heifers and cows, and further research is needed to optimize IVEP in young buffalo [30].

Regarding the impact of assisted reproductive techniques on methane emissions in cattle operations, IVEP of oocytes retrieved from young animals presents a viable approach to achieving genetic gain and reducing generation intervals [8]. Although the efficiency of IVEP in young animals is relatively lower due to hormonal and metabolic differences, its integration with genomic selection offers a powerful strategy to enhance genetic gain, efficiency, and fertility, as well as mitigate methane emissions in buffalo operations [32].

BALANCING FEED EFFICIENCY IN MEAT AND MILK PRODUCTION WITH FERTILITY AND LOW CO₂-EQ EMISSION

Ruminants are crucial in maintaining sustainable agricultural systems due to their distinctive capacity to transform forages into high-quality meat and dairy products [33]. The link between feed efficiency, methane production, and sustainability has been known for over 20 years [34, 35, 5]. The relatively high heritability of growth and feed efficiency in cattle was recognized some 70 years ago and subsequently confirmed [6, 36, 37, 38].

Furthermore, in tropical and subtropical regions, the conjunction of elevated temperatures and humidity during the summer months leads to the onset of reproductive problems, decreasing milk and meat production in buffaloes [39, 40, 41, 42]. Implementing management techniques, such as active cooling, is imperative to alleviate these stressors and uphold a certain level of productivity. Additionally, the summer season decreases feed quantity and quality, compounding the nutritional challenges that impact reproductive capabilities [39, 42]. Beyond photoperiod, it is essential to address external influences that detrimentally affect reproduction and production to fully capitalize on the potential afforded by the worldwide demand for buffalo food items. Methods encompass targeted nutritional enrichment, assisted reproductive technologies (ARTs) application, and managerial tactics (such as cooling techniques and ample resting areas) to enhance buffalo welfare within naturally endowed and non-endowed production setups.

Buffalo farming has transitioned to a more intensive model, utilizing a feeding system structured around three distinct rations corresponding to the primary buffalo production stages: lactating cows, dry cows, and growing heifers. Their diet primarily comprises maize silage and ryegrass hay, with additional concentrates reserved solely for lactating buffalo cows [43]. These farming conditions developed for buffalo production in Italy entail the absence of pasture access and wallowing water.

Recent studies suggest that incorporating more digestible forages into ruminant diets may mitigate CO₂ emissions, even within intensive systems [44]. Despite this, the cumulative emissions of free-ranging (FR) animals exceeded those of confined (C) systems by approximately 662 kg CO₂-eq. This discrepancy stemmed from the animals in the FR system consuming a greater volume of fibrous feed than the C heifers. At puberty, the heifers reached a weight of 402±3 and 382±3 kg in systems C and FR, respectively. Differences between groups were significant ($p<0.05$) due to the higher feeding regimen of group C, the higher physical activity performed while grazing by group FR, and the lower environmental temperature of the hilly area where this group was located.

Nevertheless, these animals reached puberty at an age not significantly different from that observed in group C ($p>0.05$; [45]). This finding has been attributed to the fact that grazing

animals used the available resources (pasture and feeding supplementation) efficiently. In contrast, confined heifers used spare nutrients only to increase their body mass after fulfilling their requirements for development [46].

In temperate regions, buffalo experience a distinct seasonal reproductive pattern influenced by photoperiod and melatonin secretion, as indicated by previous research [20, 47, 21]. Optimal conditions lead to a resumption of anoestrus in buffalo within 30–90 days postpartum. However, factors including inadequate nutrition and poor body condition [48], suckling management [49], and climate [50] can significantly delay this process. For instance, buffalo in Sri Lanka under free grazing with limited calf access to dams for suckling once a day resumed estrous cycles within 30–60 days, whereas those exposed to harsher conditions and free calf suckling remained in anestrus for 150–200 days [51]. Buffalo's postpartum LH secretion remains low initially, with detectable episodic pulses a few weeks before ovarian activity starts. Improved nutrition and controlled suckling prompt LH release earlier than those with poor nutrition or free suckling [52, 53]. There are recommended methods to overcome extended postpartum anestrus in buffalo, including ensuring proper nutrition before and after calving, regulating calf suckling, and alleviating heat stress through activities like wallowing or using water sprinklers [54], improving the reproductive and productive efficiency.

Limited research has been conducted on evaluating the environmental repercussions of dairy buffalo farms on environmental sustainability. In a study, Pirlo et al. [55] found that the ecological footprint of dairy buffalo farms, quantified in terms of global warming potential, amounted to 5.07 kg of CO₂ equivalent per 1 kg of standardized buffalo milk. This figure is nearly fivefold greater than that generated by dairy cow farms [56]. This disparity could be attributed to the similarity in energy inputs and raw material acquisition between dairy buffalo and cow farms, coupled with comparatively lower milk production from buffalo.

According to Chirone et al. [57], buffaloes' milk productivity varies from farm to farm and is a key factor determining environmental performance. The remaining differences are explained by a combination of the type of feed (including the portion cultivated in-house and purchased) and the strategy for managing manure. These findings reinforce the importance of increasing the genetic capacity of buffaloes to produce milk and meat more efficiently.

Buffaloes exhibit notable feed conversion efficiency and sustain productivity even when subjected to diets limiting for cattle [58]. In a previous investigation, metabolic condition and reproductive performance were observed in Murrah buffalo heifers fed either a high-energy (HE) or low-energy (LE) diet [58]. Heifers following the HE diet displayed elevated plasma levels of insulin, leptin, and T3, along with increased concentrations of IGF-1 in follicular fluid and a higher oocyte quality index. These outcomes highlight the positive effect of the nu-

trition improving the reproduction performance and production in buffaloes.

Recently, the currently used methods of estimating the carbon footprint of processed animal products and dairy products should consider the subtraction of carbon emissions and sequestration. According to De Vivo et al. [59], considering carbon sequestration and implementing this calculation method would demonstrate sustainability regarding the carbon footprint of agricultural products of animal origin, such as buffalo dairy products (Mozzarella cheese) and meat products.

ENTERIC METHANE IN PRODUCTION SYSTEM: LIFE CYCLE ASSESSMENT (LCA)

Enteric methane forms part of beef and dairy production systems' broader greenhouse gas (GHG) budget [60]. The broader GHG budget includes methane, nitrous oxide (N₂O), and CO₂ emissions from manure, feed production, vehicles and transport, and other plants and equipment. The total GHG budget of a production system is determined by life cycle assessment (LCA) methodology, standardized by ISO 14040 [61] and ISO 14044 [62] ([63, 64]).

Recently, studies were carried out to evaluate the impact of buffalo production on greenhouse gas emissions [65, 66, 57, 67].

In collaboration with Embrapa Research Institute and Cargill, our research group has studied the LCA of a buffalo milk production farm in Brazil. The data showed that enteric methane produced by buffaloes is the most relevant source of GHG production. One estimate of LCA for buffalo milk production was 63.4% for enteric methane (CH₄), 33.9% for feed production (CO₂, N₂O), and 1.92% for manure (FIG. 2).

As enteric methane has the most significant impact on the production of CO₂ equivalent, the reduction in production cycles (reduction in age at first calving and the interval between calving) and an increase in individual production (milk and meat) contribute significantly to the farm's sustainability. Furthermore, new technologies can potentially manipulate the rumen microbiome through genetic selection and various dietary intervention strategies to reduce CH₄ emissions. According to Yusuf et al. [68], methane reduction strategies have been grouped into three crucial factors: management, nutrition, and the use of advanced biotechnology. Manipulation of the rumen in reducing methane using chemicals, feed additives, roughage, concentrate utilization, and plants containing secondary compound oils has been reported [5]. Using technologies to reduce methane emissions from these crucial factors in the production chain will considerably impact the sustainability of buffalo farming).

Studies in Italy found that, despite the methane production on buffalo farms, the amount of greenhouse gases converted into CO₂-eq emitted during the buffalo dairy pro-

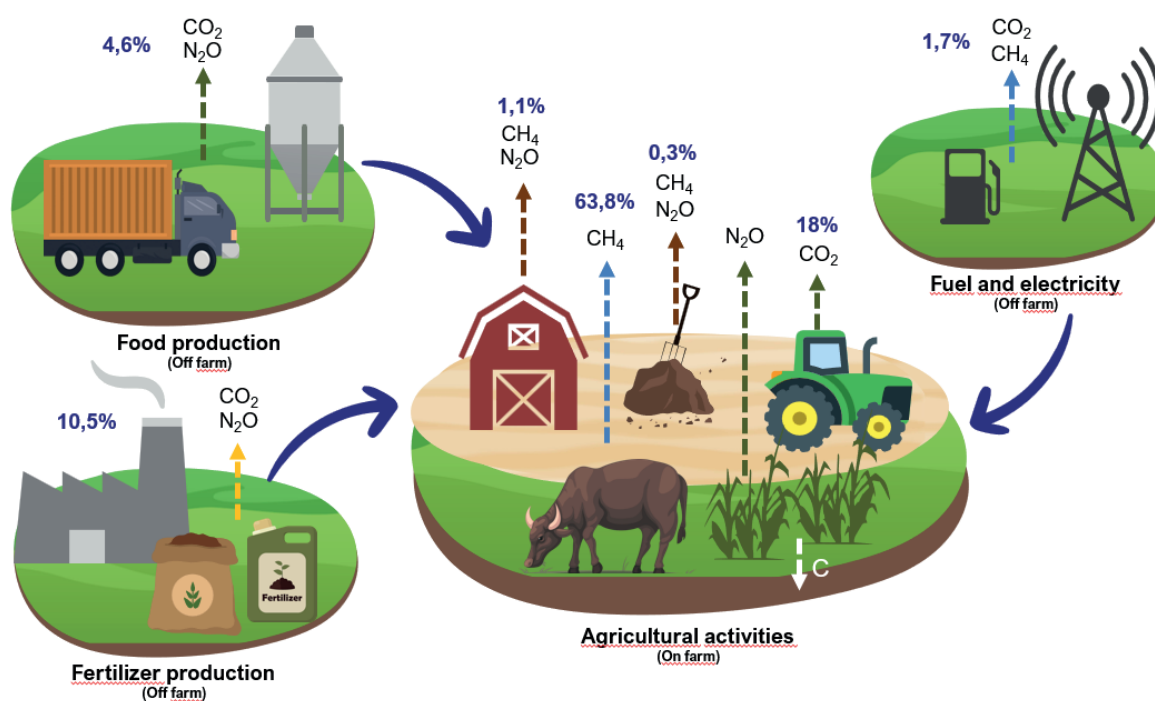


FIGURE 2. Main emission pathways related to a buffalo dairy farm in Brazil, quantified by life cycle assessment (LCA) methodology [Methodology standardized by ISO 14040 (ISO, 2006a) and ISO 14044 (ISO, 2006b); de Vries et al., 2015; Kyttä et al., 2022]. On-farm emissions represent 83.2% (agriculture activities), and off-farm emissions represent 16.8% (food production, fertilizer production, and fuel and electricity; Adapted from Abreu et al., 2023; data not yet published)

duction system is lower than the CO_2 -eq removed from the atmosphere [59]. The authors found that for every kg of buffalo Mozzarella cheese produced, 52 kg of CO_2 -eq is subtracted from the atmosphere (differences between CO_2 -eq emissions from the production system and CO_2 -eq removal from the atmosphere).

This information demonstrates that it is possible to produce beef and buffalo meat in balance with the environment, if appropriate technologies based on scientific information are used.

CONCLUSIONS AND FUTURE DIRECTION

The global attention on enteric CH_4 production in buffaloes requires a response that involves collaboration between researchers and industry. Future generations of buffaloes will be characterized by better efficiency and fertility, which may reduce CH_4 emission intensity. This will result from a balanced multi-trait selection and improved management. Artificial insemination can be incorporated into buffalo breeding programs to further improve reproductive efficiency and genetic gain, collaborating to reduce CO_2 -eq emission intensity. The urgency in moving to the next generation of buffaloes will increase the production of embryos from genomically defined prepubertal heifers. This will reduce generation interval and accelerate the rate of genetic improvement to buffaloes defined by better efficiency

and fertility and lower CH_4 emission. The growing importance of buffaloes in the world requires that they undergo an accelerated rate of genetic gain for efficiency of production, product quality, and sustainability. The challenge remains to develop integrated sustainable meat and milk production systems and communicate the importance of buffaloes for food security and the environment.

REFERENCES

- [1] United Nations Department of Economic and Social Affairs, Population Division. World Population Prospects 2022: Summary of Results. UN DESA/POP/2022/TR/NO. 3.
- [2] FAO. Sustainable Development Goals (SDG). End hunger, achieve food security, and improved nutrition, and promote sustainable agriculture. 2019. Available: <http://www.fao.org/sustainable-development-goals/news/detail-news/en/c/424259/>
- [3] Zhang Y, Colli L, Barker JSF. Asian water buffalo: Domestication, history and genetics. *Animal Genetics*. 2020; 51(2): 177-191. <https://doi.org/10.1111/age.12911>
- [4] FAO. Crops and livestock products. License: CC BY-NC-SA 3.0 IGO. Extracted from: <https://www.fao.org/faostat/en/#data/QCL>. Data of Access: 27-09-2023.

- [5] Wanapat M, Kang SC. World buffalo production: Challenges in meat and milk production, and mitigation of methane emission. *Buffalo Bulletin*. 2013, 32(1): 1–21.
- [6] Knapp Jr B, Nordskog AW. Heritability of growth and efficiency in beef cattle. *Journal of Animal Science*. 1946; 5(1): 62-70. <https://doi.org/10.2527/jas1946.5162>
- [7] Faverdin P, Guyomard H, Puillet L, Forslund A. Animal board invited review: Specialising and intensifying cattle production for better efficiency and less global warming: contrasting results for milk and meat co-production at different scales. *Animal*. 2022; 16(1): 100431. <https://doi.org/10.1016/j.animal.2021.100431>
- [8] Baruselli PS, Abreu LA, Paula VR, Carvalho B, Gricio EA, Mori FK, Rebeis LM, Albertini S, Souza AH, D’Occhio MJ. Applying assisted reproductive technology and reproductive management to reduce CO₂-equivalent emission in dairy and beef cattle: a review. *Animal Reproduction*. 2023; 20(2): e20230060. <https://doi.org/10.1590/1984-3143-AR2023-0060>
- [9] Galyean ML, Hales KE. Feeding management strategies to mitigate methane and improve production efficiency in feedlot cattle. *Animals*. 2023; 13(4): 758. <https://doi.org/10.3390/ani13040758>
- [10] Chang J, Peng S, Ciais P, Saunois M, Dangal SRS, Herrero M, Havlík P, Tian H, Bousquet P. Revisiting enteric methane emissions from domestic ruminants and their $\delta^{13}\text{CCH}_4$ source signature. *Nature Communications*. 2019; 10: 3420. <https://doi.org/10.1038/s41467-019-11066-3>
- [11] FAO. Reducing enteric methane for improving food security and livelihoods. 2021. Available: <http://www.fao.org/in-action/enteric-methane/background/why-is-enteric-methane-important/en/>
- [12] Bowen JM, Cormican P, Lister SJ, McCabe MS, Duthie CA, Roehe R, Dewhurst RJ. Links between the rumen microbiota, methane emissions and feed efficiency of finishing steers offered dietary lipid and nitrate supplementation. *PLoS One*. 2020; 15(4): e0231759. <https://doi.org/10.1371/journal.pone.0231759>
- [13] Ross EM, Moate PJ, Maret L, Cocks BG, Hayes BJ. Investigating the effect of two methane-mitigating diets on the rumen microbiome using massively parallel sequencing. *Journal of Dairy Science*. 2013; 96(9): 6030-6046. <https://doi.org/10.3168/jds.2013-6766>
- [14] Knapp JR, Laur GL, Vadas PA, Weiss WP, Tricarico JM. Invited review: Enteric methane in dairy cattle production: Quantifying the opportunities and impact of reducing emissions. *Journal of Dairy Science*. 2014; 97(6): 3231-3261. <https://doi.org/10.3168/jds.2013-7234>
- [15] Roehe R, Dewhurst RJ, Duthie CA, Rooke JA, McKain N, Ross DW, Hyslop JJ, Waterhouse A, Freeman TC, Watson M, Wallace RJ. Bovine host genetic variation influences rumen microbial methane production with best selection criterion for low methane emitting and efficiently feed converting hosts based on metagenomic gene abundance. *PLoS Genetics*. 2016; 12(2): e1005846. <https://doi.org/10.1371/journal.pgen.1005846>
- [16] Baruselli PS, Carvalho NAT, Gasparrini B, Campanile G, D’Occhio MJ. Review: Development, adoption, and impact of assisted reproduction in domestic buffaloes. *Animal*. 2023; 17(S1): 100764. <https://doi.org/10.1016/j.animal.2023.100764>
- [17] Malik PK, Trivedi S, Mohapatra A, Kolte AP, Sejian V, Bhatta R, Rahman H. Comparison of enteric methane yield and diversity of ruminal methanogens in cattle and buffaloes fed on the same diet. *PLoS ONE*. 2021; 16(8): e0256048. <https://doi.org/10.1371/journal.pone.0256048>
- [18] Baruselli, PS, Carvalho JGS, Elliff FM, Da Silva JCB, Chello D, Carvalho NAT. Embryo transfer in buffalo (*Bubalus bubalis*). *Theriogenology*. 2020; 150: 221-228. <https://doi.org/10.1016/j.theriogenology.2020.01.037>
- [19] Hegarty RS, McEwan JC. Genetic opportunities to reduce enteric methane emissions from ruminant livestock. In ‘Proceedings of the Ninth World Congress on Genetics Applied to Livestock Production’; 2010; Leipzig, Germany. Leipzig: German Society for Animal Science. 2010. pp 181-186.
- [20] Baruselli PS, Carvalho NAT, Gimenes LU, Crepaldi GA. Fixed-time artificial insemination in buffalo. *Italian Journal of Animal Science*. 2007; 6(S2): 107-118. <https://doi.org/10.4081/ijas.2007.s2.107>
- [21] Zicarelli L. Reproductive seasonality in buffalo. *Bubalus Bubalis*. 1997; 4(29): 52–54.
- [22] D’Occhio MJ, Ghuman SS, Neglia G, Della Valle G, Baruselli PS, Zicarelli L, Visintin JA, Sarkar M, Campanile G. Exogenous and endogenous factors in seasonality of reproduction in buffalo: A review. *Theriogenology*. 2020; 150: 186-192. <https://doi.org/10.1016/j.theriogenology.2020.01.044>
- [23] Zicarelli L. Can we consider buffalo a non precocious and hypofertile species? *Italian Journal of Animal Science*. 2007; 6(S2): 143-154. <https://doi.org/10.4081/ijas.2007.s2.143>
- [24] Carvalho NAT, Soares JG, Porto Filho RM, Gimenes LU, Souza DC, Nichi M, Sales JS, Baruselli PS. Equine chorionic gonadotropin improves the efficacy of a timed artificial insemination protocol in buffalo during the non-breeding season. *Theriogenology*. 2013; 79(3): 423-428. <https://doi.org/10.1016/j.theriogenology.2012.10.013>
- [25] Monteiro BM, Souza DC, Vasconcellos GSFM, Carvalho NAT, Baruselli PS. Effect of season on dairy buffalo reproductive performance when using P4/E2/eCG-based fixed-time artificial insemination management. *Therio-*

- genology. 2018; 119: 275-281. <https://doi.org/10.1016/j.theriogenology.2018.07.004>
- [26] Baruselli PS, Madureira EH, Barnabe VH, Barnabe RC, Berber RCA. Evaluation of synchronization of ovulation for fixed timed insemination in buffalo (*Bubalus bubalis*). *Brazilian Journal of Veterinary Research and Animal Science*. 2003; 40(6): 431-442. <https://doi.org/10.1590/S1413-95962003000600007>
- [27] Sá Filho MF, Carvalho NAT, Gimenes LU, Torres-Júnior JR, Nasser LFT, Tonhati H, Garcia JM, Gasparini B, Zicarelli L, Baruselli, PS. Effect of recombinant bovine somatotropin (bST) on follicular population and on in vitro buffalo embryo production. *Animal Reproduction Science*. 2009; 113(1-4): 51-59. <https://doi.org/10.1016/j.anireprosci.2008.06.008>
- [28] Carvalho JGS, Carvalho NAT, Bayeux BM, Watanabe YF, Watanabe OY, Mingoti RD, Baruselli PS. Superstimulation prior to the ovum pick-up improves the in vitro embryo production in nulliparous, primiparous and multiparous buffalo (*Bubalus bubalis*) donors. *Theriogenology*. 2019; 138: 164-168. <https://doi.org/10.1016/j.theriogenology.2019.07.003>
- [29] Silva JCB, Rezende RG, Colli MHA, Bayeux, BM, Mingoti RD, Ojeda-Rojas OA, Basso AC, Naves JR, Baruselli PS. In vitro embryo production in buffalo: Comparison between calves, prepubertal heifers and lactating cows. *Animal Reproduction*. 2017; 14(3): 766.
- [30] Baldassarre H, Bordignon V. Laparoscopic ovum pick-up for in vitro embryo production from dairy bovine and buffalo calves. *Animal Reproduction*. 2018; 15(3): 191-196. <https://doi.org/10.21451/1984-3143-AR2018-0057>
- [31] Baruselli PS, Soares JG, Bayeux BM, Da Silva JCB, Mingoti RD, Carvalho NAT. Assisted reproductive technologies (ART) in water buffaloes. *Animal Reproduction*. 2018; 15(1): 971-983. <http://dx.doi.org/10.21451/1984-3143-AR2018-0043>
- [32] Currin L, Baldassarre H, Bordignon V. In vitro production of embryos from prepubertal Holstein cattle and Mediterranean water buffalo: Problems, progress and potential. *Animals*. 2021; 11(8): 2275. <https://doi.org/10.3390/ani11082275>
- [33] Baruselli PS, Rodrigues CA, Ferreira RM, Sales JNS, Elliff FM, Silva LG, Viziack MP, Factor L, D'Occhio MJ. Impact of oocyte donor age and breed on in vitro embryo production in cattle, and relationship of dairy and beef embryo recipients on pregnancy and the subsequent performance of offspring: A review. *Reproduction, Fertility and Development*. 2021; 34(2): 36-51. <https://doi.org/10.1071/RD21285>
- [34] Cammack KM, Austin KJ, Lamberson WR, Conant GC, Cunningham HC. Ruminant Nutrition Symposium: Tiny but mighty: The role of the rumen microbes in livestock production. *Journal of Animal Science*. 2018; 96(2): 752-770. <https://doi.org/10.1093/jas/skx053>
- [35] Johnson DE, Ward GM. Estimates of animal methane emissions. *Environmental Monitoring and Assessment*. 1996; 42: 133-141. <https://doi.org/10.1007/BF00394046>
- [36] Arthur PF, Herd RM. Efficiency of feed utilisation by livestock - Implications and benefits of genetic improvement. *Canadian Journal of Animal Science*. 2005; 85(3): 281-290. <https://doi.org/10.4141/A04-062>
- [37] Berry DP, Crowley JJ. Cell Biology Symposium: Genetics of feed efficiency in dairy and beef cattle. *Journal of Animal Science*. 2013; 91(4): 1594-1613. <https://doi.org/10.2527/jas2012-5862>
- [38] Gonzalez-Recio O, Pryce JE, Haile-Mariam M, Hayes BJ. Incorporating heifer feed efficiency in the Australian selection index using genomic selection. *Journal of Dairy Science*. 2014; 97(6): 3883-3893. <https://doi.org/10.3168/jds.2013-7515>
- [39] Sypniewski M, Strabel T, Pszczola M. Genetic variability of methane production and concentration measured in the breath of Polish Holstein-Friesian cattle. *Animals*. 2021; 11(11): 3175. <https://doi.org/10.3390/ani11113175>
- [40] Perera BMAO. Reproductive cycles of buffalo. *Animal Reproduction Science*. 2011; 124(3-4): 194-199. <https://doi.org/10.1016/j.anireprosci.2010.08.022>
- [41] Khan FA, Das GK, Pande M, Sarkar M, Mahapatra RK, Shankar U. Alterations in follicular fluid estradiol, progesterone and insulin concentrations during ovarian acyclicity in water buffalo (*Bubalus bubalis*). *Animal Reproduction Science*. 2012; 130(1-2): 27-32. <https://doi.org/10.1016/j.anireprosci.2011.12.020>
- [42] Abdoon AS, Gabler C, Holder C, Kandil OM, Einspanier R. Seasonal variations in developmental competence and relative abundance of gene transcripts in buffalo (*Bubalus bubalis*) oocytes. *Theriogenology*. 2014; 82(8): 1055-1067. <https://doi.org/10.1016/j.theriogenology.2014.07.008>
- [43] Phogat JB, Pandey AK, Singh I. Seasonality in buffaloes reproduction. *International Journal of Plant and Animal Sciences*. 2016; 6(2): 46-54.
- [44] Sabia E, Napolitano F, Claps S, Braghieri A, Piazzolla, N, Pacelli C. Feeding, nutrition and sustainability in dairy enterprises: the case of Mediterranean buffaloes (*Bubalus bubalis*). In: Vastola, A. (Ed.), *The Sustainability of Agro-food and Natural Resource Systems in the Mediterranean Basin*. 2015. Springer Open, pp. 57-64. https://doi.org/10.1007/978-3-319-16357-4_5
- [45] Sabia E, Claps S, Napolitano F, Annicchiarico G, Bruno A, Francaviglia R, Sepe L, Aleandri R. In vivo digestibility

- of two different forage species inoculated with arbuscular mycorrhiza in Mediterranean red goats. *Small Ruminant Research*. 2015; 123: 83-87. <https://doi.org/10.1016/j.smallrumres.2014.10.008>
- [46] Sabia E, Napolitano F, Salvatore C, De Rosa G, Barile VL, Braghieri A, Pacelli C. Environmental impact of dairy buffalo heifers kept on pasture or in confinement. *Agricultural Systems*. 2018; 159: 42-49. <https://doi.org/10.1016/j.agsy.2017.10.010>
- [47] Sabia E, Napolitano F, De Rosa G, Terzano GM, Barile VL, Braghieri A, Pacelli C. Efficiency to reach age of puberty and behaviour of buffalo heifers (*Bubalus bubalis*) kept on pasture or in confinement. *Animal*. 2014; 8(11): 1907-1916. <https://doi.org/10.1017/S1751731114001876>
- [48] Borghese A. Buffalo Production and Research. Technical Series 67. 2005. Rome, Italy: Food and Agriculture Organization of the United Nations.
- [49] Baruselli OS, Barnabe VH, Barnabe RC, Visintin JA, Molero-Filho JR, Porto R. Effect of body condition score at calving on postpartum reproductive performance in buffalo. *Buffalo Journal*. 2001; 1: 53-65.
- [50] Usmani RH, Dailey RA, Inskeep EK. Effects of limited suckling and varying prepartum nutrition on postpartum reproductive traits of milked buffaloes. *Journal of Dairy Science*. 1990; 73(6): 1564-1570. [https://doi.org/10.3168/jds.S0022-0302\(90\)78826-1](https://doi.org/10.3168/jds.S0022-0302(90)78826-1)
- [51] Nanda AS, Brar PS, Prabhakar S. Enhancing reproductive performance in dairy buffalo: Major constraints and achievements. *Reproduction Supplement*. 2003; 61: 27-36.
- [52] Perera BMAO, de Silva LNA, Kuruwita VY, Karunaratne AM. Postpartum ovarian activity, uterine involution and fertility in indigenous buffaloes at a selected village location in Sri Lanka. *Animal Reproduction Science*. 1987; 14(2): 115-127. [https://doi.org/10.1016/0378-4320\(87\)90091-1](https://doi.org/10.1016/0378-4320(87)90091-1)
- [53] Mohan V, Kuruwita VY, Perera BMAO, Abeygunawardena H. Effects of suckling on the resumption of postpartum ovarian activity in buffaloes. *Tropical Agricultural Research*. 1990; 2: 306-315.
- [54] Singh AK, Brar PS, Nanda AS, Prakash BS. Effect of suckling on basal and GnRH-induced LH release in postpartum dairy buffaloes. *Animal Reproduction Science*. 2006; 95(3-4): 244-250. <https://doi.org/10.1016/j.anireprosci.2005.10.004>
- [55] Perera BMAO, Abeygunawardena H, Vale WG, Chantalakhana C. Buffalo. In: *Livestock and Wealth Creation – Improving the Husbandry of Animals Kept by Poor People in Developing Countries*. Livestock Production Programme. Natural Resources International Limited. UK. 2005; pp 451–471.
- [56] Pirlo G, Caré S, Fantin V, Falconi F, Buttol P, Terzano GM, Masoni P, Pacelli C. Factors affecting life cycle assessment of milk produced on 6 Mediterranean buffalo farms. *Journal of Dairy Science*. 2014; 97(10): 6583-6593. <http://dx.doi.org/10.3168/jds.2014-8007>
- [57] O'Brien D, Shalloo L, Patton J, Buckley F, Grainger C, Wallace M. A life cycle assessment of seasonal grass-based and confinement dairy farms. *Agricultural Systems*. 2012; 107: 33-46. <https://doi.org/10.1016/j.agsy.2011.11.004>
- [58] Chirone R, Paulillo A, Salatino P, Salzano A, Cristofaro B, Cristiano T, Campanile G, Neglia G. Life cycle assessment of buffalo milk: A case study of three farms in southern Italy. *Journal of Cleaner Production*. 2022; 365: 132816. <https://doi.org/10.1016/j.jclepro.2022.132816>
- [59] Campanile G, Baruselli PS, Vecchio D, Prandi A, Neglia G, Carvalho NAT, Sales JNS, Gasparrini B, D'Occhio MJ. Growth, metabolic status and ovarian function in buffalo (*Bubalus bubalis*) heifers fed a low energy or high energy diet. *Animal Reproduction Science*. 2010; 22(1-2): 74-81. <https://doi.org/10.1016/j.anireprosci.2010.07.005>
- [60] De Vivo R, Zicarelli L, Napolano R, Zicarelli F. Calculation method of the carbon footprint of products of animal origin integrated with the physiological absorption of carbon dioxide: Calculation example of the CFP of mozzarella di Bufala Campana DPO. *Advances in Environmental and Engineering Research*. 2023; 4(3): 044. <https://doi.org/10.21926/aeer.2303044>
- [61] Ibdhi R, Calsamiglia S. Carbon footprint assessment of Spanish dairy cattle farms: Effectiveness of dietary and farm management practices as a mitigation strategy. *Animals*. 2020; 10(11): 2083. <https://doi.org/10.3390/ani10112083>
- [62] ISO 14040. Environmental management - life cycle assessment - principles and framework. 2006. Reference number ISO 14040:2006(E).
- [63] ISO 14044. Environmental management - life cycle assessment - requirements and guidelines. 2006. Reference number ISO 14044:2006(E).
- [64] de Vries M, van Middlelaar CE, de Boer IJM. Comparing environmental impacts of beef production systems: A review of life cycle assessments. *Livestock Science*. 2015; 178: 279-288. <https://doi.org/10.1016/j.livsci.2015.06.020>
- [65] Kyttä V, Roitto M, Aastapsev A, Saarinen M, Tuomisto HL. Review and expert survey of allocation methods used in life cycle assessment of milk and beef. *The International Journal of Life Cycle Assessment*. 2022; 27: 191-204. <https://doi.org/10.1007/s11367-021-02019-4>
- [66] Ijaz M, Goheer MA. Emission profile of Pakistan's agriculture: past trends and future projections. *Environment*,

- Development and Sustainability. 2021; 23: 1668-1687. <https://doi.org/10.1007/s10668-020-00645-w>
- [67] Romano E, De Palo P, Tidona F, Maggiolino A, Bragaglio A. Dairy buffalo Life Cycle Assessment (LCA) affected by a management choice: The production of wheat crop. Sustainability. 2021; 13(19): 11108. <https://doi.org/10.3390/su131911108>
- [68] Correddu F, Lunesu MF, Caratzu MF, Pulina G. Recalculating the global warming impact of italian livestock methane emissions with new metrics. Italian Journal of Animal Science. 2023; 22(1): 125-135. <https://doi.org/10.1080/1828051X.2023.2167616>
- [69] Yusuf RO, Noor ZZ, Abba AH, Abu Hassan MA, Mohd Din MF. Greenhouse gas emissions: Quantifying methane emissions from livestock. American Journal of Engineering and Applied Sciences. 2012; 5(1): 1-8. <https://doi.org/10.3844/ajeassp.2012.1.8>